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Synthesis of a Chiral 2,6-Bridged Morpholine System: *trans*-6,7-Diol Derivatives of 8-Oxa-3-azabicyclo[3.2.1]octane

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Abstract: The title compound 6, [1R-(6endo,7exo)]-8-oxa-3-azabicyclo[3.2.1]octane-6,7-diol, was derived in nine steps from 1-amino-1-deoxy-D-glucitol 2. The key step was the acid catalyzed rearrangement of epoxide 3 to the 2,5-cis-disposed hydrofuran compound 5. Linkage of the 2,5-substituents, to form the bridged secondary amine 23, proceeded via mesylation of 6-OH and deprotection of 1-NH₂. Further O-debenzylation and (or) N-substitution afforded the aminodiol 6 and various N-alkylated derivatives. On basis of ¹H NMR analysis the chair form was assigned to the bridged morpholine ring.

INTRODUCTION

Recently we reported the synthesis of various 7-carbonyl homologues of 1-deoxynojirimycin 1, i.e the acid, amide and ketone compounds. These were derived from 1-amino-1-deoxy-D-glucitol 2 via a crystalline 3,4;5,6-diacetonide salt. In a concurrent sequence, 6-azido and 6-amino analogues of 1 were prepared via the N-Boc-2,3;5,6-di-O-isopropylidene derivative of aminoalditol 2. The present work deals with the conversion of 2 to the epoxide 3, which might serve as an alternative precursor of the azasugars 1 (1-deoxynojirimycin) and 4 (castanospermine) and their analogues. In non aqueous acidic medium, epoxide 3 rearranges to the cis-hydrofuran compound 5 which can be transformed to the chiral bridged aminodiol 6.

The non chiral parent compound 8-oxa-3-azabicyclo[3.2.1]octane 7 was prepared in 1948.⁴ The N-substituted derivatives 8 and 9 exhibit analgesic and antiinflamatory activities in mice and rats.⁵ The related 3-azabicyclo[3.2.1]octane compound 10 acts as an analgesic in rats.⁶ Bicyclic compounds 11 (X = heteroatom) with specific groups attached to X or incorporated into R display potent antiarrhythmic activity in animal models.⁷ The herewith reported compound 6 can be viewed as a chiral analogue of the bicyclic structures 7, 10, and 11.

RESULTS AND DISCUSSION

We commenced our synthesis with the aminoalditol 2 which is utilized as a precursor of the antidiabetic compound 1 and some N-substituted analogues. The amino group of 2 was protected as the N-trityl derivative 12 (Scheme 1). The latter was prepared via initial silylation of the OH groups, N-tritylation, and deprotection of the trimethylsilyl groups with K2CO3 in methanol, in 71% overall yield. Compound 2 is not soluble in non aqueous medium. It has been reported that tritylation of amino acids in aqueous medium gives low yields of N-tritylamino acids due to hydrolysis of trityl chloride. Whereas these difficulties can be resolved by applying the above-mentioned silyl protection method, we also developed an alternative procedure to prepare compound 12 directly from the p-toluenesulfonate salt of 2. In contrast to the free amine 2, this salt is soluble in pyridine. Sequential addition of Et3N and (C6H5)3CCl afforded 12 in 81% yield.

Scheme 1. Reagents: (a) 1) HMDS, TMSCl, pyridine, CH₂Cl₂, reflux, 3 h; 2) TrCl, Et₃N, CH₂Cl₂, 0 °C, 4 h; 3) K₂CO₃, MeOH, 4 h; (b) 1) TsOH (1 equiv.), MeOH; 2) Pyridine, Et₃N, TrCl; (c) 1) (MeO)₃CH, PPTS, THF, 50°C, 1-3 h; 2) NaH, DMF, BnBr, 0 °C, 1 h; (d) PPTS, aq.MeOH, THF; (e) MsCl, DMAP, Et₃N, CH₂Cl₂, room temp.; (f) MeONa, MeOH; (g) TsOH.H₂O or Me₃SO₃H, THF, room temp., 10 min.

To prepare epoxide 3, selective protection of the 5- and 6-OH groups was required. This was accomplished by heating 12 with trimethyl orthoformate and no more than 0.04 equivalents of pyridinium p-toluenesulfonate (PPTS) in THF at 50 °C for 1 to 3 hours. Prolonged reaction times or a higher concentration of PPTS led to less polar side products. After benzylation of the crude product with benzyl bromide and NaH in DMF, the resulting compound 13 was partially hydrolysed with aqueous methanol and PPTS to afford the pure 6-O-formyl ester 14 in 51% overall yield from 12. None of the 5-O-formyl regioisomer was detected in the ¹H NMR spectrum. Mesylation of 14 in dichloromethane with MsCl, DMAP, and Et₃N followed by treatment with MeONa in MeOH, produced an inseparable 9:1 mixture of the epoxides 3 and 17 in 97% yield. This mixture was used in the next steps. Presumably, the formation of epoxide 17 from 14 proceeds via the 6-O-mesylate 16 which results from the base (Et₃N) catalysed

migration of the 6-O-formyl group of 14 to the 5-O-position and mesylation of the primary hydroxyl group. The N-benzyl-N-trifluoroacetyl analogue of epoxide 3 was used previously in the synthesis of 1-deoxynojirimycin and castanospermine.³

Treatment of the mixture of epoxides 3 and 17 with p-toluenesulfonic acid or methanesulfonic acid in THF afforded the five-membered ring compounds 5 (main product) and 18 (trace component). The mass spectra of these compounds were very similar, showing the presence of the N-trityl group and only two benzyl groups: ions corresponding to [M-C₆H₅]⁺ and [M-Tr]⁺ were detected at m/z 508 and 342 respectively. Opening of the epoxide and generation of an OH group was indicated by the introduction of one trimethylsilyl group (mass shift of 72 Daltons). In the 400 MHz ¹H NMR spectrum of compound 5 in d_6 -DMSO, the primary alcohol proton was detected as a triplet centered at δ 4.70 ppm (J 6 Hz). The $1\overset{\circ}{3}$ C NMR spectrum revealed the disappearance of signals corresponding to the original epoxide 3 (δ 42.7 for C-6 and δ 53.5 ppm for C-5), and confirmed the presence of two benzyl groups. On the basis of these results and the coupling constant values observed in the ¹H NMR spectrum, the five-membered ring structure 5 was attributed to the main product. The cis relationship for C-1 and C-6, which implies a double inversion at C-5 (steps f and g, Scheme 1), was verified by transformation of the main product 5 to the bridged compound 23 (see below). Presumably, attack of the 2-OBn oxygen at C-5 of the protonated epoxide results in a S_N2-type cyclization and debenzylation, as depicted (Scheme 2) for both the major and the minor epoxide compounds. Similar cyclisations to hydrofuran compounds, resulting from participation of ether oxygens, have been reported previously. 11 When the mixture of epoxides 3 and 17 was heated in acidic aqueous methanol, the resulting compounds 20a and 20b were found to be the same (TLC, mass spectra) as those obtained indirectly via N-detritylation of 5 and 18. The 3,4,6-tri-O-benzyl analogue of cis compound 20a has been reported as a 5:1 mixture with the C-2 epimer. 12

Linkage of the *cis*-disposed 2,5-substituents of compound 5 provides access to a chiral bridged morpholine system in which chirality relates to the *trans* orientation of identical substituents on the bridge carbons 6 and 7 (Scheme 3). The desired transformation $(5 \rightarrow 23)$ was accomplished *via* mesylation of 6-OH, acid deprotection of 1-NH₂, and heating of the primary amine salt 22.HCl with triethylamine in dichloromethane. The cyclization could be followed through CIMS analysis of the *N*-acetylated reagent and product components (22, MH⁺ 464 and 23, MH⁺ 368). Following chromatographic separation as the free base 23, partial degradation was prevented by acidifying the eluates with HCl prior to evaporation. The crystalline HCl-salt was isolated in 57% yield calculated on 5.

Difficulties were encountered in debenzylation of compound 23. Even at 50-60 °C, only one benzyl group was removed on hydrogenation of the HCl-salt with Pd/C in methanol-acetic acid (CIMS: MH⁺ 234). However, smooth debenzylation of both protecting groups was achieved by using Pearlman's Pd(OH)₂ catalyst with HCl in methanol. Chromatographic isolation as the free base afforded the crystalline aminodiol 6 in 94% yield.

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Scheme 3. Reagents: (a) MsCl, DMAP, Et₃N, CH₂Cl₂, room temp.; (b) TsOH.H₂O or MeSO₃H, MeOH, 60 °C, 30 min; (c) CH₂Cl₂, Et₃N, reflux 30 min; (d) MeOH, HCl, Pd(OH)₂, H₂, room temp.; (e) MeOH, CH₂O, 10% Pd/C, H₂; (f) MeOH, Et₃N, 28, reflux, 2 h.

To characterize further the bicyclic system and to assess the biologic activity of representative compounds, we prepared some N-alkyl and O-deprotected analogues. The N-methyl compounds 24 and 25 were derived via hydrogenation of 23 and 6 with formaldehyde. Reaction of 23 with the racemic epoxide 28 in methanol at reflux temperature gave a mixture of epimeric compounds 26a,b. Both the epimeric mixture and the chromatographically purified isomers were hydrogenated with Pd(OH)₂, to afford the corresponding aminodiols 27a,b. N-Substitution of various amino compounds with the 3-(2-or 4-fluorophenoxy)-2-hydroxypropyl group gave rise to products showing diverse pharmacological profiles. ¹³⁻¹⁵

Analysis of the 400 MHz ¹H NMR spectra (Table 1) clearly showed the chair conformation of the bridged morpholine ring. This form was indicated by similar ³J-values (1.8-2.6 Hz) measured for coupling between the bridgehead protons H-1 and H-5 and the axial and equatorial neighbouring protons H-2 and H-4. The ensuing nearly identical torsional angles (ca. 60°) preclude alternative boat or flattened chair conformations. Supporting evidence for the chair conformation of the morpholine ring comes from a comparison with the structurally related cocaine isomers 29a-d. For all four isomers, attribution of the chair form to the ethano bridged piperidine ring likewise rested on similar values for ³J_{1,2} (2.2-3.3 Hz), ³J_{4eq,5} (2.2-3.1 Hz), and ³J_{4ax,5} (1.8-4.8 Hz). ¹⁶ A crystallographic study of the related ketone compound 30 also showed a chair-like conformation for the six-membered pyran ring. ¹⁷

In the spectrum of the N-methyl compound 25, equatorial endo protons H-2 and H-4 were differentiated by a characteristic long-range coupling (1.8 Hz). Decoupling of the low-field proton H-4endo removed this 4J -coupling and also reduced the signal multiplicity for the bridgehead proton H-5 (dt \rightarrow dd). The remaining dd signal relates to coupling with the cis-oriented protons H-6 (6.5 Hz) and H-4exo (2 Hz). In contrast to H-6exo, H-7endo displayed a rather sharp singlet ($\omega \frac{1}{2} = 2$ Hz) indicating torsional angles of ca. 90° with the trans-oriented protons H-6 and H-1. These data are comparable to those reported for norbornane-type compounds. ¹⁸

In the C_6D_6 spectrum of compound 24, signals for the bridgehead protons H-1, H-5 coincided (δ 4.1), as shown by the NOE's induced on irradiation of either H-2 or H-4. To assign the latter protons, we performed ^{13}C -{ ^{1}H } selective decoupling of H-1,H-5. This resulted in singlet signals for C-1 and C-5, and in residual couplings for C-2 and C-4 (triplet and dd, respectively) which correlated with the separations observed for each pair of protons (H-2: δ 0.1 ppm; H-4: δ 0.6 ppm). The carbons C-2 and C-4 in turn were

identified on basis of the characteristic (anti- and syn-) γ -effects¹⁹ originating from the exo and endo benzyloxy groups (C-2: δ 58.3; C-4: δ 53.6). Finally, signals for the bridgehead protons were separated (δ 0.02 ppm) by applying a solvent mixture C₆D₆-CDCl₃; further ¹H-¹H decoupling confirmed the assignment of vicinal protons H-1, H-2 and H-4, H-5.

	23.HCl	24	25
Proton	δ (ppm), multiplicity, J (Hz)	δ (ppm), multiplicity, J (Hz)	δ (ppm), multiplicity, J (Hz)
H-2 _{exo}	3.02, d (br), 12	2.24, dd, 11.2, 1.8	1.93, dd, 11,2.5
H-4 _{exo}	2.83, d (br), 12	2.14, dd, 11.6, 2.6	1.98, dd, 11.5, 2
H-2 _{endo}	3.52, d (br), 12	2.35, d (br), 11	2.08, dt, 11, 1.8, 1.8
H-4 _{endo}	2.98, d (br), 12	2.77, d (br), 11.6	2.50, dt 11.5, 1.8, 1.8
H-1	3.88, s (br)	4.10, m	3.73, s, $\omega_{\frac{1}{2}}$: 5.5
H-7 _{endo}	5.12, s (br)	4.37, s (br)	3.89, s, $\omega_{\frac{1}{2}}$: 2
H-5	3.72, d (br)	4.10, m	4.07, dt, 6.5, 2, 1.8
H-6 _{exo}	4.17, d, 6	4.26, dd, 6.2, 1.4	4.23, d, 6.5 Hz
CH ₃	-	2.06, s	1.67, s
CH ₂ -Ar	4.39, 4.43, ABq, 11.5	4.38, 4.45, ABq, 11 Hz	1.07, \$
CH ₂ -Ar	4.38, 5.05, ABq , 11.5	4.37, s	-
Ar-H	7.20-7.40, m	7.08-7.40, m	-

Table 1. 400 MHz 1H NMR of compounds 23, 24, and 25 in C_6D_6

In the ¹H NMR spectra of the secondary amines **6** and **23**, signals for the bridge and bridgehead protons H-1, H-5, H-6, H-7 were similar to those described for the *N*-methyl compounds (see Experimental). Due to overlap and broadening effects, however, no definite assignments could be made with regard to H-2 and H-4. For the HCl-salt of **23**, some broadening of signals also was observed. The assignments shown in Table 1 were based on decoupling of H-6 and a low-field proton H-2 (tentatively assigned as H-2*endo*); NOE performed for H-2, H-4, and H-5 enhanced H-1, H-5, and H-6, respectively.

In preliminary in vitro experiments using various receptor systems, no significant receptor affinity was observed for the bridged morpholino compounds 6 and 23-27.

CONCLUSION

Starting from 1-amino-1-deoxy-D-glucitol, we developed a synthetic route to a 2,6-bridged morpholine system. In these bicyclic products, chiral positions 3,4 and 2,5 of D-glucose are transposed as the bridge and bridgehead carbons 6, 7 and 5, 1 respectively. The morpholine ring, comprising positions 2, 3, and 4 as the mobile portion, was shown to be in a chair conformation implying the *endo* orientation for the amino nitrogen. In addition to the synthetic and conformational aspects discussed so far, substitution of the hydroxyl and amino groups of aminodiol 6 allows for attachment of spatially directed groups, e.g. pharmacophores and fluorophores.

EXPERIMENTAL SECTION

General methods

Melting points were uncorrected. The optical rotations were measured on a Propol polarimeter fitted with a 7 cm cell. IR spectra were recorded as thin films between NaCl plates on a Perkin-Elmer 297 grating IR spectrophotometer. ¹H and ¹³C NMR were recorded on a Bruker AMX 400 instrument operating at 400 MHz for ¹H and 100 MHz for ¹³C. ¹H and ¹³C chemical shifts are reported in ppm relative to tetramethylsilane as an internal reference. *J* values are reported in Hz. Mass spectra were run on Kratos MS50 and Hewlett-Packard 5989A instruments; the ion source temperature was 150-250°C as required. Exact masses were measured at a resolution of 10,000. Analytical and preparative thin layer chromatography were performed using Merck silica gel 60 PF-224. Column chromatography was carried out using 70-230 mesh silica gel 60 (E. M. Merck). Dry solvents were freshly distilled. Solutions were dried over MgSO₄. 1-Amino-1-deoxy-D-glucitol was supplied by Cerestar. Elemental analysis was performed by Janssen Pharmaceutica on a Carbo Erla elemental analyser type 1106.

1-Amino-1-deoxy-1-N-triphenylmethyl-p-glucitol (12).

Method A: Trimethylsilyl chloride (40 ml, 77 mmol) and 1,1,1,3,3,3-hexamethyldisilazane (20 ml, 94 mmol) were added to a suspension of 2 (2.20 g, 11.3 mmol) in pyridine (10 ml) and CH₂Cl₂ (10 ml). The mixture was heated under reflux for 3 h. After cooling, the excess of reagents was destroyed by slow addition of MeOH. The solvent was removed, the oily residue was redissolved in CH₂Cl₂ and the solution washed with cold water. To the dried and cooled (0 °C) solution, TrCl (3.60 g, 12.1 mmol) and Et₃N (5 ml) were added and the mixture stirred for 4 h at 0 °C. After washing with water and evaporation of CH₂Cl₂, the residue was dissolved in MeOH (15 ml) containing K₂CO₃ (5 g). The desilylation was complete after 4 h. The mixture was evaporated to dryness and the residue distributed between CH₂Cl₂ and water. The CH₂Cl₂ phase was dried and concentrated to a small volume. This was added to an excess of hexanes and the precipitated solid collected by filtration. After three precipitations in hexanes as above, pure 12 was obtained in 71% (3.41 g) overall yield: mp 125-126°C; [α]_D^{18.5}-9.8° (c 1, MeOH); ¹H NMR (400 MHz, CDCl₃) δ 2.35 (m, 2 H), 3.53 (m, 2 H), 3.70 (m, 4 H), 7.15-7.45 (m, 15 H); ¹³C NMR (100 MHz, CDCl₃), δ 46.6 (CH₂), 63.9 (CH₂), 70.8 (C), 71.7 (CH), 72.0 (CH), 72.3 (CH), 73.4 (CH), 126.6-128.7 (aromatic CH), 145.2 (aromatic C).

Method B: To a stirred suspension of 2 (11.4 g, 60 mmol) in MeOH (150 ml) was added p-toluenesulfonic acid monohydrate (11.4 g, 60 mmol). After 20 min the mixture became homogeneous. The solvent was removed by co-evaporation with toluene and CCl₄, respectively. The crystalline residue was redissolved in pyridine (20 ml). Dry Et₃N (50 ml) was added followed immediately by TrCl (17 g, 60.4 mmol). After 30 min, another portion of TrCl (6 g, 21.3 mmol) was added and the reaction was allowed to proceed for 1 h. The solvent was removed and the residue was distributed between CH₂Cl₂ and water. After workup as above, compound 12 was isolated in 81% yield (20.5 g).

1-Amino-2,3,4-tri-O-benzyl-1-deoxy-6-O-formyl-1-N-triphenylmethyl-D-glucitol (14). Compound 12 (6.20 g, 14.6 mmol) was heated at 50 °C with (MeO)₃CH (4.80 ml, 44 mmol) and PPTS (0.070 g, 0.28 mmol) in THF (25 ml) for 1.5 h. The mixture was allowed to reach room temp., and an aqueous solution of K₂CO₃ was added. After evaporation of THF, the residue was distributed between CH₂Cl₂ and water. The organic phase was dried and evaporated. The residue was dissolved in DMF (20 ml) and the solution added to a cooled (0 °C) mixture of NaH (80% dispersed in mineral oil, 2.20 g, 73 mmol) and DMF (20 ml). After 10 min of stirring, benzyl bromide (7.1 ml, 59 mmol) was added. After completion of the reaction (2 h), the excess of reagent was destroyed by slow addition of MeOH (5 ml). The solution was diluted with

CH₂Cl₂ (60 ml) and the solution washed thoroughly with water. The CH₂Cl₂ layer was dried and evaporated to afford an oily residue containing 13 as the major product. Without purification, this residue was dissolved in 100 ml of a mixture of THF/MeOH/H₂O (10/9/1). PPTS (3.17 g, 14.7 mmol) was added and the mixture stirred for 6 h. After evaporation of the solvent and column chromatography (EtOAc/hexanes: 2:3), 14 (5.4 g) was isolated as an oil in 51% overall yield from 12. $[\alpha]_D^{20}$ -0.65° (c 3.5, MeOH); IR v_{max} (cm⁻¹): 3435, 3035, 2927, 1725, 1597, 1494, 1452, 1088; ¹H NMR (400 MHz, CDCl₃) δ 2.25 (m, 1 H, H-1a), 2.47 (m, 1 H, H-1b), 3.58 (dd, J = 8, 4 Hz, 1 H, H-4), 3.90 (m, 2 H, H-2, H-3), 4.00 (m, 1 H, H-5), 4.18 (dd, 2J = 12, 3J = 6 Hz, 1 H, H-6a), 4.35 (dd, 2J = 12, 3J = 3 Hz, 1 H, H-6b), 4.37, 4.45 (ABq, 2J = 11.5 Hz, 2 H, CH₂Ph, 4.49 (s, 2 H, CH₂Ph), 4.65, 4.72 (ABq, 2J = 11.5 Hz, 2 H, CH₂Ph), 6.96-7.54 (m, 30 H, 6 Ph), 8.07 (s, 1 H, HCO-); 13C NMR (100 MHz, CDCl₃) δ 43.4 (C-1), 65.2 (C-6), 69.7 (C-5), 70.7 (Ph₃C), 72.9, 73.2, 74.0 (3 CH₂Ph), 76.7 (C-4), 78.7, 79.2 (C-2, C-3), 126.3, 127.8, 127.8, 128.0, 128.1, 128.4, 128.5, 128.6 (aromatic CH), 137.4, 137.6, 137.9 (C-1' CH₂Ph), 145.7 (C-1' Ph₃C), 160.9 (HCO); HRMS calcd for C₄1H₄2NO₆ ([M-Ph]⁺) 644.3012, found 644.3010, calcd for C₂8H₃2NO₆ ([M-Tr]⁺) 478.2230, found 478.2237.

1-Amino-5,6-anhydro-2,3,4,-tri-O-benzyl-1-deoxy-1-N-triphenylmethyl-L-iditol (3). To a solution of 14 (2.20 g, 3.0 mmol) in dry CH₂Cl₂ (50 ml) were added MsCl (0.36 ml, 4.5 mmol), DMAP (0.372 g, 3.0 mmol), and Et3N (0.70 ml, 9.6 mmol) at room temp. The mixture was stirred for 1 h after which time it was washed with water (3x50 ml). The organic layer was dried and evaporated to dryness. The residue was dissolved in MeOH (15 ml), MeONa (0.329 g, 6.1 mmol) was added and the solution was stirred for 2 h. Water (35 ml) was added and the solution extracted with dichloromethane (3x50 ml). The dichloromethane layer was evaporated and the residue purified by column chromatography (EtOAc/hexanes: 3:17) to afford 2.94 g (97%) of an oil which consisted of an inseparable 9:1 mixture of epoxides 3 and 17. ¹H NMR (400 MHz, CDCl₃) 3: δ 2.02 (dd, 2J = 13, 3J = 5 Hz, 1 H, H-1a), 2.28 (dd, 2J = 5, $^3J_{trans}$ = 2.5 Hz, 1 H, H-6a), 2.35 (dd, $^2J = 13$, $^3J = 4$ Hz, 1 H, H-1b), 2.40 (t, $^2J = 5$, $^3J_{cis} = 4$ Hz, 1 H, H-6b), 3.01 (dd, J = 7, 3 Hz, 1 H, H-4), 3.12 (ddd, J = 7, 4, 2.5 Hz, 1 H, H-5), 3.83 (dd, J = 7, 3 Hz, 1 H, H-3),3.92 (m, 1 H, H-2), 4.33, 4.47, 4.52, 4.64, 4.73, 4.78 (6 H, 3 CH_2Ph), 7.20-7.40 (39 H, Ph). 17: δ 2.61 (dd, 2J = 5 Hz, $^3J_{trans}$ = 2.5 Hz, 1 H, H-6b), 2.68 (dd, 2J = 5 Hz, $^3J_{cis}$ = 3.5 Hz, 1 H, H-6a), 3.10 (ddd, $^{3}J = 5$ Hz, $^{3}J_{cis} = 3.5$ Hz, $^{3}J_{trans} = 2.5$ Hz, 1 H, H-5), 3.28 (dd, J = 5, 2.5 Hz, 1 H, H-4). 3: ¹³C NMR (100 MHz, CDCl₃) δ 42.7 (C-6), 42.8 (C-1), 53.5 (C-5), 70.4 (CPh₃), 71.9, 72.8, 74.5 (OCH₂Ph), 79.6, 79.9, 80.2 (C-2, C-3, C-4), 126.2-145.8 (aromatic carbons). HRMS calcd for C₄₀H₄₀NO₄ ([M-Ph]⁺) 598.2957, found 598.2922, calcd for C₂₇H₃₀NO₄ ([M-Tr]⁺) 432.2175, found 432.2176.

1-Amino-2,5-anhydro-3,4-di-*O*-benzyl-1-deoxy-1-*N*-triphenylmethyl-D-glucitol (5). To a solution of the 9:1 mixture of 3 and 17 (0.94 g, 1.4 mmol) in THF (20 ml) was added *p*-TsOH.H₂O (0.845 g, 4.4 mmol). After 10 min, the mixture was made alkaline with aq.K₂CO₃ and extracted with CH₂Cl₂. Column chromatography (hexanes/EtOAc: 7:3) afforded 5 (0.44 g, 53%) as an oil. [α]_D²⁰ +18.8° (*c* 4.59, CHCl₃); IR: v_{max} (cm⁻¹) 3426, 3085, 3060, 3030, 2872, 1596, 1176, 1099; ¹H NMR (400 MHz, CDCl₃) δ 2.45 (m, 2 H, 2 H-1), 3.61 (dd, ²*J* = 11.5, ³*J* = 4 Hz, 1 H, H-6a), 3.78 (dd, ²*J* = 11.5, ³*J* = 3 Hz, 1 H, H-6b), 4.02 (dd, *J* = 4.5, 1.5 Hz, 1 H, H-3), 4.05-4.09 (m, *J* = 3, 1.5 Hz, 2 H, H-4, H-5), 4.28 (td, *J* = 6, 4.5 Hz, 1 H, H-2), 4.29, 4.54 (ABq, ²*J* = 12 Hz, 2 H, CH₂Ph), 7.00-7.50 (m, 26 H, 5 *Ph*). ¹³C NMR (100 MHz, CDCl₃) δ 43.2 (CH₂), 63.1 (CH₂), 70.8 (C),

71.3 (CH₂), 72.0 (CH₂), 81.0 (CH), 82.7 (CH), 82.8 (CH), 84.1 (CH), 126.2, 127.6, 127.7, 127.8, 127.9, 128.0, 128.6, (aromatic CH), 137.2 (C), 137.6 (C), 145.9 (3 C); HRMS calcd for C₃₉H₃₉NO₄ (M⁺) 585.2879, found 585.2858. The more polar isomer 18 was isolated as a trace component by preparative TLC (hexanes/EtOAc: 7:3) and was characterized *via* mass spectral comparison with compound 5. The isomeric compounds 5 and 18 were analysed both as the free alcohols and trimethylsilyl derivatives prepared by heating with *N*,*O*-bis(trimethylsilyl)trifluoroacetamide in pyridine.

[1R-(6endo,7exo)]-6,7-di-O-Benzyl-8-oxa-3-azabicyclo[3,2.1]octane-6,7-diol (23). To a solution of 5 (15.0 g, 25.6 mmol) in dry CH₂Cl₂ (100 ml) was added MsCl (3.20 ml, 40 mmol), DMAP (1.60 g, 13 mmol), and Et₃N (12 ml) at room temp. The mixture was stirred for 15 min after which time it was washed with water (3x50 ml). The organic layer was dried and evaporated to furnish crude 21. Without purification, the oily residue was dissolved in 100 ml of a 1:1 mixture of concentrated HCl and MeOH. After being heated at 60 °C for 30 min, the solution was evaporated to dryness. A mixture of the crude salt 22.HCl and Et₃N (20 ml) in CH₂Cl₂ (100 ml), was heated under reflux for 30 min (after 1 min and 30 min, respectively, samples of the solution were treated with Ac₂O for CIMS analysis). The solvent was removed and the residue purified by column chromatography (MeOH/CHCl3: 3:47). To prevent partial degradation, fractions containing the free base 23 were acidified with methanolic HCl. Evaporation of the solvent afforded 5.32 g (57%) of crystalline 23.HCl. 23: oil, $[\alpha]_{D}^{20}$ -6.4° (c 0.6, CHCl₃); IR ν_{max} (cm⁻¹): 3085, 3040, 2963, 2869, 1497; ¹H NMR (400 MHz, C₆D₆) δ 2.40 (d, ²J = 13 Hz, 1 H), 2.80 $(d, {}^{2}J = 13 \text{ Hz}, 1 \text{ H}), 2.95 (d, {}^{2}J = 13 \text{ Hz}, 1 \text{ H}), 2.98 (d, {}^{2}J = 13 \text{ Hz}, 1 \text{ H}), 3.91 (br s, 1 H, H-1), 4.01$ (br s, 1 H, H-5), 4.09 (d, J = 2 Hz, 1 H, H-7endo), 4.21 (dd, J = 6, 2 Hz, 1 H, H-6exo), 4.23, 4.37 $(ABq, {}^{2}J = 12 Hz, 2 H, CH_{2}Ph), 4.29, 4.31 (ABq, {}^{2}J = 12 Hz, 2 H, CH_{2}Ph), 7.10-7.40 (m, 15 H, Ph);$ ¹³C NMR (100 MHz, CDCl₃) δ 45.1 (CH₂) , 48.3 (CH₂), 71.3 (CH₂), 72.9 (CH₂), 76.7 (CH), 79.2 (CH), 86.0 (CH), 86.7 (CH), 127.8, 127.9, 128.4, (aromatic CH), 137.7 (aromatic C); HRMS calcd for C₂₀H₂₃NO₃ (M⁺) 325.1688, found 325.1678; 23.HCl, mp 60 °C; ¹H NMR (Table 1). Anal. calcd. for C₂₀H₂₄ClNO₃.H₂O: C, 63.24; H, 6.90; N, 3.69; O, 16.85; H₂O, 4.74. Found C, 63.43; H, 6.77; N, 3.59; O, 16.10; H₂O, 4.17.

[1R-(6endo,7exo)]-8-Oxa-3-azabicyclo[3.2.1]octane-6,7-diol (6). To a solution of 23.HCl (0.500 g, 1.5 mmol) in MeOH (30 ml), were added 5 drops of concentrated HCl and Pd(OH)₂ (0.125 g). The mixture was hydrogenated in a Parr apparatus (35 psi) for 3 h. The catalyst was removed by filtration through Celite, the filtrate was evaporated and the residue purified by column chromatography on silica gel (H₂O/NH₄OH/MeOH/CHCl₃: 1:1:28:70) to afford 0.188 g (94%) of the crystalline aminodiol 6: mp 195-196.6 °C; $[\alpha]_D^{20}$ -4.4° (c 2.3, MeOH); ¹H NMR (400 MHz, CD₃OD), δ 3.18-3.38 (m, 4 H, H-2, H-4), 4.07 (br s, 1 H, H-1), 4.20 (br s, 1 H, H-7endo), 4.39 (br d, J = 7 Hz, 1 H, H-6exo), 4.49 (br d, J = 7 Hz, 1 H, H-5); ¹³C NMR (100 MHz, CD₃OD) δ (ppm) 44.4 (CH₂), 45.8 (CH₂), 76.2 (CH), 80.5 (2 CH), 81.7 (CH); HRMS calcd for C₆H₁₁NO₃ (M⁺) 145.0740, found 145.0746.

[1R-(6endo,7exo)]-6,7-Di-O-benzyl-3-methyl-8-oxa-3-azabicyclo[3.2.1]octane-6,7-diol (24). To a solution of 23.HCl (0.47 g, 1.4 mmol) in MeOH (30 ml), were added an aqueous solution (37%) of formaldehyde (2 ml) and 10% Pd/C (0.114 g). The mixture was hydrogenated in a Parr apparatus (35 psi) for 1 h. The catalyst was removed by filtration through Celite and the filtrate was made alkaline with methanolic ammonia. After concentration, the residue was dissolved in MeOH and the methanol solution was evaporated. This treatment was repeated several times in order to convert a more polar complex to compound 24. The residue was dissolved in CH₂Cl₂ and the resulting solution washed with water. The organic phase was dried, and evaporated. The residue was purified by column chromatography (hexanes/EtOAc: 7:3) to

give 0.427 g (97%) of the N-methyl compound 24 as an oily residue: $[\alpha]_D^{20}+2.2^\circ$ (c 0.96, CHCl₃); IR: ν_{max} (cm⁻¹) 3100-3090, 2960-2850, 1550, 1500; ¹H NMR (Table 1); ¹³C NMR (100 MHz, C₆D₆) δ 45.6 (CH₃), 53.9 (C-4), 58.3 (C-2), 71.6 (CH₂Ph), 72.6 (CH₂Ph), 75.6, 79.1 (C-1, C-5), 86.4 (C-7), 87.4 (C-6), 127.8-128.5 (aromatic CH), 139.00 (aromatic C); HRMS calcd for C₂₁H₂₅NO₃ (M⁺) 339.1834, found 339.1826.

[1R-(6endo,7exo)]-3-Methyl-8-oxa-3-azabicyclo[3.2.1]octane-6,7-diol (25). To a solution of 23.HCl (0.400 g, 1.1 mmol) in MeOH (30 ml), were added 5 drops of concentrated HCl and Pd(OH)₂ (0.100 g). The mixture was hydrogenated in a Parr apparatus (35 psi) for 3 h. The catalyst was removed by filtration through Celite. The filtrate was evaporated and to the residue were added MeOH (30 ml), paraformaldehyde (0.13 g, 4.3 mmol) and 10% Pd/C (0.100 g). The mixture was hydrogenated in a Parr apparatus (35 psi) for 3 h. The catalyst was removed by filtration through Celite, the filtrate was evaporated and the residue was chromatographed on a silica gel column (H₂O/NH₄OH/MeOH/CHCl₃: 1:1:28:70) to afford 0.083 g (47% yield) of the crystalline aminodiol 25: mp 126.6°, $[\alpha]_D^{20}$ +35.2° (c 0.6, CHCl₃); IR: ν_{max} (cm⁻¹) 3550-3400, 2960-2850; ¹H NMR (Table 1); ¹³C NMR (100 MHz, C₆D₆) δ (ppm) 45.0 (CH₃), 55.4 (CH₂), 57.5 (CH₂), 76.8 (CH), 81.1 (CH), 82.6 (CH), 84.0 (CH); HRMS calcd for C₇H₁₃NO₃ (M⁺) 159.0895, found 159.0902.

1-{[1R-(6endo,7exo)]-6,7-Di-O-benzyl-6,7-dihydroxy-8-oxa-3-azabicyclo[3.2.1]oct-3-yl}-3-p-

fluorophenoxy-2-propanol (26a and 26b). EtaN (1 ml) and racemic epoxide 28 (0.665g, 4.0 mmol) were added to a solution of 23.HCl (0.992 g, 2.74 mmol) in MeOH (40 ml). The solution was heated under reflux for 30 min. The solvent was removed and the residue was purified by column chromatography (EtOAc/CHCl3: 3:47). Three fractions were isolated (98% combined yield): these consisted of the pure compounds 26a (29%, less polar) and 26b (25%, more polar), and a mixed fraction 26a,b. 26a: oil, $[\alpha]_D^{20}$ -16.2° (c 1.01, CHCl₃); IR: v_{max} (cm⁻¹) 3440, 3100-3090, 2960-2850, 1580, 1500; ¹H NMR (400 MHz, C₆D₆) δ 2.13 (br d, 1 H), 2.36-2.40 (m, 3 H), 2.56 (br d, 1 H), 2.79 (br d, 1 H), 3.75 (dd, J = 10, 4.5 Hz, 1 H), 3.85 (dd, J = 10, 5.4 Hz, 1 H), 4.04 (m, 3 H), 4.18 (br s, 1 H), 4.22 (dd, J = 7,1.5 Hz, 1 H), 4.29 (s, 2 H), 4.41 (s, 2 H), 6.70-6.90 (m, 4 H), 7.20-7.40 (m, 10 H); ¹³C NMR (100 MHz, C₆D₆) δ 50.4 (CH₂), 57.4 (CH₂), 59.7 (CH₂), 65.5 (CH), 71.2 (CH₂), 71.4 (CH₂) 73.6 (CH₂), 75.6 (CH), 78.9 (CH), 87.5 (CH), 87.8 (CH), 115.8-115.9 (aromatic CH), 127.7-128.7 (aromatic CH), 137.8, 138.7, 155.5, 158.9 (aromatic C); HRMS calcd for C₂₉H₃₂NO₅F (M⁺) 493.2264, found 439.2263; 26b: oil, $[\alpha]_D^{20}$ +10.2° (c 1.61, CHCl₃); IR: v_{max} (cm⁻¹) 3440, 3100-3090, 2960-2850, 1580, 1500; ¹H NMR $(400 \text{ MHz}, C_6D_6) \delta 2.27 \text{ (br d, 1 H)}, 2.40 \text{ (m, 4 H)}, 2.63 \text{ (br d, 1 H)}, 3.69 \text{ (dd, } J = 9, 5 \text{ Hz, 1 H)}, 3.83$ (dd, J = 9, 5 Hz, 1 H), 3.90 (br s, 1 H), 4.04 (s, 2 H, CH₂), 4.09 (br s, 1 H), 4.15 (br d, 1 H), 4.22-4.38 (m, 4 H), 6.65-6.90 (m, 4 H), 7.20-7.40 (m, 10 H); ¹³C NMR (100 MHz, C₆D₆) δ 53.4 (CH₂), 55.0 (CH₂), 59.9 (CH₂), 66.4 (CH), 70.9 (CH₂), 71.3 (CH₂), 73.2 (CH₂), 75.8 (CH), 78.6 (CH), 86.9 (CH), 87.6 (CH), 115.7-115.9 (aromatic CH), 127.7-128.7 (aromatic CH), 137.9, 138.4, 155.3, 158.7 (aromatic C); HRMS calcd for C₂₉H₃₂NO₅F (M⁺) 493.2264, found 493.2262.

1-{[1R-(6endo,7exo)]-6,7-Dihydroxy-8-oxa-3-azabicyclo[3.2.1]oct-3-yl}-3-p-fluorophenoxy-2-propanol (27b). To a solution of 26b (0.200 g, 0.455 mmol) in MeOH (30 ml), were added 5 drops of concentrated HCl and Pd(OH)₂ (0.050 g). The mixture was hydrogenated in a Parr apparatus (35 psi) for 3 h. The catalyst was removed by filtration through Celite, the filtrate was evaporated and the residue was purified by column chromatography (MeOH/CHCl₃: 1:9) to afford 0.114 g (80%) of compound 27b as an oil: $[\alpha]_D^{20}$

+16.0° (c 3.2, CHCl₃); IR: $ν_{\rm max}$ (cm⁻¹) 3550-3450, 3100-3090, 2960-2850, 1560; ¹H NMR (400 MHz, CDCl₃) δ 2.49 (m, 3 H), 2.63 (d, 1 H), 2.87 (d, 2 H), 3.82 (m, 2 H), 4.03 (m, 2 H), 4.19-4.32 (m, 3 H), 6.75-7.00 (m, 4 H); ¹³C NMR (100 MHz, CDCl₃) δ 54.0 (CH₂), 54.4 (CH₂), 59.6 (CH₂), 66.7 (CH), 71.1 (CH₂), 77.2 (CH), 81.4 (CH), 81.8 (CH), 82.5 (CH), 115.6-116.1 (aromatic CH), 154.6, 158.7 (aromatic C); HRMS calcd for C₁₅H₂₀NO₅F (M⁺) 313.1325, found 313.1321.

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